

Photosynthetic abilities, light response, and stomatal function in six agroforestry species, *Dipterocarpus tuberculatus*, *D. alatus*, *Eucalyptus camaldulensis*, *Hevea brasiliensis*, *Colocasia gigantea*, and *C. esculenta* in responses to water deficit

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ABSTRACT: We investigated the physiological and morphological responses in six agroforestry tree species *Dipterocarpus tuberculatus*, *D. alatus*, *Eucalyptus camaldulensis*, *Hevea brasiliensis*, *Colocasia gigantea*, *C. esculenta* under limited water availability. Withholding water resulted in reduced field capacity by 71%, 86%, and 48% in *D. alatus*, *D. tuberculatus*, and *E. camaldulensis*, respectively, resulting in wilting, leaf chlorosis, and leaf burn, respectively. Stomatal density varied among the plant species (1.4–6.2 stomata per 0.0625 mm²) and was unaffected by water regimes, whereas stomatal conductance (g_s), transpiration rate (E), and water use efficiency were reduced when plants were subjected to water deficit conditions. Generally, intracellular CO₂ of well-watered plants was not always maintained across increasing light intensity flux (PAR_i), although for *E. camaldulensis* this was not the case, as both well-watered and water deficit groups showed a reduction with increasing PAR_i. Net photosynthetic rate was significantly increased in well-watered plants as compared to water stressed plants, depending on the degree of PAR_i. In addition, a positive relation between g_s and E in each plant was demonstrated.

KEYWORDS: drought situation, photosynthetic photon flux, evapotranspiration rate, water efficiency

INTRODUCTION

Agroforestry is an initiative for promoting environmental rehabilitation in domestic community through the integrated management of trees rather than mono-plantations^{1,2}. A sustainable land use system, intercropping forest trees, and agricultural/horticultural crops is a challenge. It can help in sequestration of atmospheric carbon into the soil², supplementing food products³, improving their nutritional status⁴, increasing the density of soil arbuscular mycorrhizal fungi⁵, elevating crop productivity⁶, and thus providing increased yield, food security, and economic income⁷. The physiological and morphological responses, specifically photosynthesis and biomass production, in millet (cereal

crop), taro (storage root crop) and baobab (forest tree) intercropping system have been well established, allowing the coexistence of canopy and understorey plant species despite competitive shading for better biomass production⁶. Shading and water availability in the root zones influence the biomass production, and are considered as the major factors in controlling the agroforestry system^{8–10}. Hence the light response curve of plant species under water limitation plays a key factor in agroforestry management and needs to be investigated.

Global warming and climate change have resulted in increased drought stress and is currently a matter of concern¹¹. In deciduous agroforests, water constraint in the dry season is evidently found in each year. Stomatal function under water deficit

conditions that prevent the water loss via stomatal closure has been investigated. In contrast, it limits the CO₂ assimilation and plant growth^{12,13}. Drought sensitivity in higher plants depends on the degree of water limitation, plant species, drought exposure periods, their interactions, etc^{14,15}. Plant species show individual responses to water-deficit stress as mentioned in large number of previous publications. Hence the basic knowledge on physiological, morphological and anatomical characters in plant responses to water shortage, especially in the summer and winter season, especially in agroforest tree species are restricted. Stomatal density in olive cultivars increases whereas the stomatal size is reduced, relating to drought stress (−1.5 MPa soil water potential) for 7 months¹⁶. Stomatal conductance is one of the most important parameters to identify the stomata closure, which is very sensitive to drought conditions¹⁷. The stomatal function in dynamic process of CO₂ assimilation and H₂O transpiration has been validated in terms of water use efficiency, generally declined when plants subjected to water-deficit stress¹⁸. Overall growth inhibition, leaf chlorosis (chlorophyll degradation), leaf burn and leaf falling of tropical species in summer and winter seasons are evidently observed¹⁹. Sensitivity of plant species to water shortage in the dry season (water shortage) has been well established in terms of visual symptoms, i.e., survival percentage²⁰, relative mortality rate²¹, drought index²² and wilting state²³. Furthermore, the agroforestry design relating to light requirements and their adaptive abilities to water limited conditions is still lacking. The aim of this study was to assess differences in physiology in responses to varying water availability and light intensity for the establishment of novel agroforestry systems, relating to individual requirements of plant species.

MATERIALS AND METHODS

Plant materials and treatments

Seeds of *Dipterocarpus alatus* and *D. tuberculatus* were collected from the mixed deciduous forest in the northern region of Thailand, and germinated in plastic bags containing mixed soil (electrical conductivity 2.687 dS/m; pH 5.5; organic matter 10%; total nitrogen 0.17%; total phosphorus 0.07%; total potassium 1%). One year-old *Hevea brasiliensis* scion cv. RRIM 600 on wild root stock was cultivated in clay pot culture and *Eucalyptus camaldulensis* plantlets derived from micropropagation²⁴ were transplanted and grown in a greenhouse under 500–

1000 μmol m^{−2} s^{−1} photosynthetic photon flux density (PPFD) with a 10 h per day photoperiod, temperature 28 ± 2 °C, and 80 ± 5% relative humidity (RH). Corms of *Colocasia esculenta* and *C. gigantea* were collected from swamp habitat in the northern region of Thailand. The plants were divided into two groups: (a) well watering (WW) (2 times daily irrigation) and (b) water deficit (WD) condition (water withholding for 7 d) with 5 plants per group. Field capacity (FC, %) was measured and calculated by weighing the plastic bag containing mixed soil at 0, 2, 4, and 7 d after water withholding. Intracellular CO₂ (C_i), net photosynthetic rate (P_n), stomatal conductance (g_s), transpiration rate (E), water use efficiency (WUE) and stomatal density in the leaf tissues were measured. All plants were grown in a greenhouse under 500–1000 μmol m^{−2} s^{−1} PPFD with a 10 h per day photoperiod, 28 ± 2 °C and 80 ± 5% RH in the plastic bag containing mixed soil and incubated in a greenhouse for four months.

Photosynthetic, stomatal function, and WUE

P_n, g_s, E, and WUE of the second fully matured leaf from the shoot tip was measured by a portable photosynthesis system connected to an infra-red gas analyser (Model LI 6400XT, LI-COR Inc., Lincoln, NE, USA). The air-flow rate of sample chamber was 500 μmol CO₂ s^{−1} at 25 °C and the light intensity was 1000 μmol m^{−2} s^{−1} PPFD of 6400-02B red-blue LED light source. Light response curve in each plant species was derived based on auto-measurement at 0, 50, 100, 250, 500, 750, 1000, and 1500 μmol m^{−2} s^{−1} (PPFD or PAR_i). WUE was calculated according to a ratio of P_n and E.

Stomatal density assay

Clear nail polish was painted on the abaxial surface of the leaf, until dry, then peeled from the leaf using Scotch Tape (transparent type, Scotch) subsequently mounted peel in a grid (0.0625 mm²) under a compound 400× light microscope (model Axiostarplus, Carl Zeiss NY, USA) connected with a Motic microscope camera (Motic, Hong Kong) and the number of stomata observed under 5 fields of each peel was recorded²⁵.

Experiment design and statistical analysis

The experiment was arranged in completely randomized design with five replicates (n = 5). The mean values obtained in each plant under well watering and water-deficit stress were compared using *t*-test and analysed with SPSS (version 11.5).

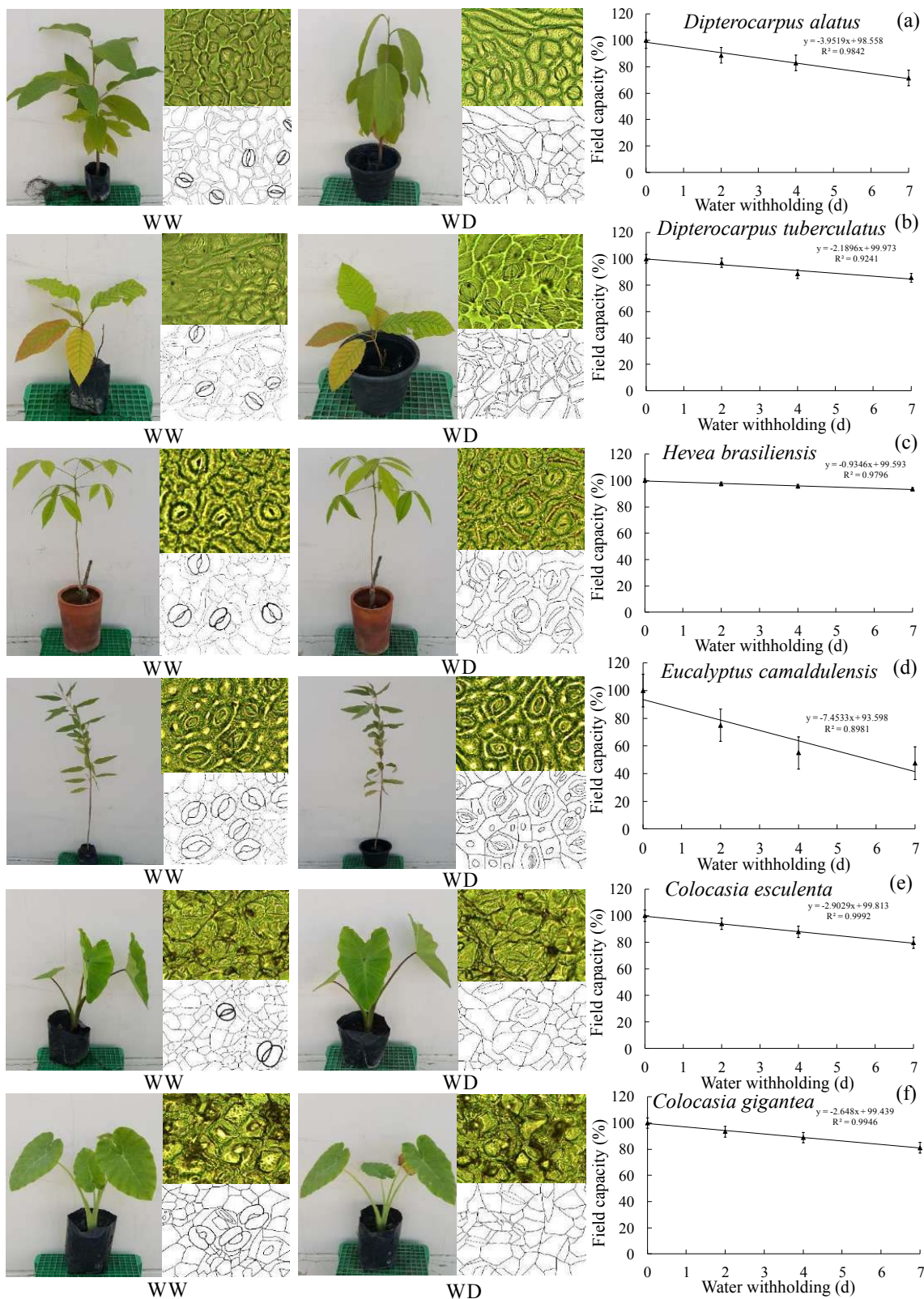


Fig. 1 Morphological characteristics, stomatal density in the plant grown under well watering (WW) and decline field capacity (%) in the soil substrate after water withholding for 7 days.

Table 1 Net photosynthetic rate (P_n), stomatal conductance (g_s), transpiration rate (E), water use efficiency (WUE), and stomata density in *D. alatus* Roxb. ex G. Don, *D. tuberculatus* Roxb., *H. brasiliensis* Muell. Arg., *E. camaldulensis* Dehnh., *C. esculenta* (L.) Schott, and *C. gigantea* Hook. f. grown under well watering (WW) and water deficit (WD) conditions. Standard errors were expressed as mean \pm SE.

Plant species	Water	P_n	g_s	E	WUE	Stomatal density
<i>D. alatus</i>	WW	2.91 \pm 0.19 ^a	0.051 \pm 0.002 ^a	1.48 \pm 0.06 ^a	19.8 \pm 1.4 ^a	6.00 \pm 0.84
	WD	0.34 \pm 0.03 ^b (88%)	0.016 \pm 0.001 ^b (69%)	0.30 \pm 0.02 ^b (80%)	11.6 \pm 1.0 ^b (42%)	5.80 \pm 0.97
<i>D. tuberculatus</i>	WW	4.88 \pm 0.27 ^a	0.064 \pm 0.004 ^a	1.40 \pm 0.08 ^a	34.7 \pm 3.0 ^a	6.20 \pm 0.58
	WD	0.89 \pm 0.05 ^b (82%)	0.020 \pm 0.003 ^b (69%)	0.57 \pm 0.08 ^b (22%)	15.8 \pm 1.3 ^b (54%)	5.40 \pm 0.68
<i>H. brasiliensis</i>	WW	2.49 \pm 0.10 ^a	0.057 \pm 0.001 ^a	1.10 \pm 0.03 ^a	22.5 \pm 0.4 ^a	4.60 \pm 0.51
	WD	1.69 \pm 0.13 ^b (32%)	0.010 \pm 0.001 ^b (82%)	0.86 \pm 0.02 ^b (22%)	19.7 \pm 1.1 ^b (12%)	3.60 \pm 0.51
<i>E. camaldulensis</i>	WW	6.60 \pm 0.33 ^a	0.083 \pm 0.005 ^a	2.72 \pm 0.12 ^a	24.5 \pm 1.6 ^a	6.00 \pm 0.63
	WD	0.71 \pm 0.04 ^b (89%)	0.007 \pm 0.001 ^b (92%)	0.42 \pm 0.03 ^b (85%)	17.2 \pm 0.6 ^b (30%)	5.80 \pm 0.49
<i>C. esculenta</i>	WW	6.97 \pm 0.23 ^a	0.102 \pm 0.002 ^a	2.25 \pm 0.04 ^a	31.1 \pm 1.2 ^a	2.20 \pm 0.37
	WD	3.39 \pm 0.05 ^b (51%)	0.038 \pm 0.003 ^b (63%)	1.40 \pm 0.06 ^b (38%)	24.3 \pm 1.0 ^b (22%)	1.60 \pm 0.40
<i>C. gigantea</i>	WW	3.55 \pm 0.15 ^a	0.042 \pm 0.004 ^a	2.25 \pm 0.04 ^a	31.1 \pm 1.2 ^a	2.00 \pm 0.32
	WD	1.68 \pm 0.12 ^b (53%)	0.014 \pm 0.001 ^b (67%)	1.40 \pm 0.06 ^b (38%)	24.3 \pm 1.0 ^b (22%)	1.40 \pm 0.25

Different letters in each column show significant difference at $p \leq 0.01$ by t -test. Parentheses represent reduction percentage of water-deficit stressed plants compare to well watering in each parameter.

RESULTS

Morphological and physiological responses to WD condition

Field capacity (%) of the soil decreased depending on the duration water withholding. After 7 d water withholding period, FC sharply declined in the plastic bag containing *E. camaldulensis* plant (48% FC), with leaf burn symptoms. Furthermore, 71% FC was measured in the soil containing *D. alatus*, with initiation of wilting symptoms. In contrast, FC measured in the soil of *C. gigantea*, *C. esculenta*, *D. tuberculatus* and *H. brasiliensis* were 80%, 85%, 86%, and 93%, respectively, without any morphological changes after 7 d water withholding (Fig. 1). There was no significant difference in stomatal density among treatments (Table 1), whereas stomatal closure in all plant species under water deficit condition was observed (Fig. 1).

In the present study, P_n , g_s , E , and WUE were sensitive to a degree of FC reduction in the soil (Table 1). In *E. camaldulensis*, P_n , g_s , E , and WUE in plants under 48% FC were dropped by 89%, 92%, 85%, and 30%, respectively, when compared with well watering. Likewise, these parameters in

D. alatus were declined by 88%, 69%, 80%, and 42%, respectively, when subjected to 71% FC. P_n , g_s , and WUE in *D. tuberculatus* under 86% FC were diminished by 82%, 69%, and 54%, respectively. Interestingly, these parameters in *Colocasia* species were slightly decreased when exposed to 85% FC (Table 1). In *H. brasiliensis*, the FC maintained at 93% resulted in high P_n , E , and WUE.

Light response curves under WD condition

In emergent layers of *Dipterocarpus* spp., intracellular CO_2 ($\mu\text{mol CO}_2 \text{ mol}^{-1}$) was declined, causing an increased input of photosynthetically active radiations (PAR_i). C_i in well-watered plants of *D. alatus* was higher than that in water deficit condition, while it was unchanged in *D. tuberculatus* (Fig. 2a,b). CO_2 assimilation rate or P_n in both *D. alatus* and *D. tuberculatus* was increased along with the degree of light intensities (Fig. 2c,d), especially in well-watered plants. *D. tuberculatus* was sensitive to light intensity (Fig. 2d). In water deficit condition, it was slightly increased when compared with well watering (Fig. 2c,d). Furthermore, a positive relation between g_s and E in *D. alatus* ($R^2 = 0.987$) and *D. tuberculatus* ($R^2 = 0.921$) was

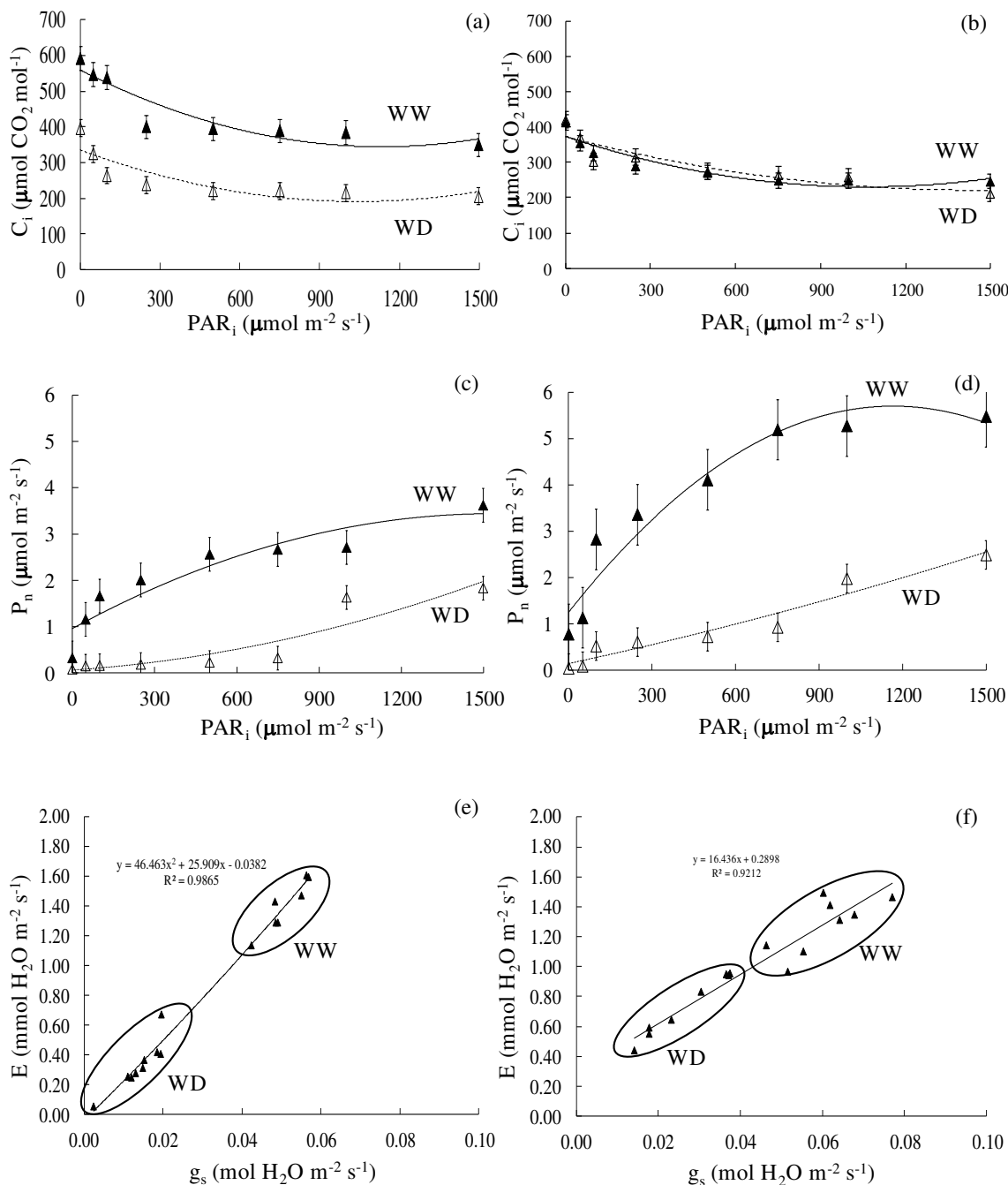


Fig. 2 Response of (a,b) intracellular CO₂ (C_i) and (c,d) net photosynthetic rate (P_n) to light intensity flux (PAR_i) and (e,f) relationship between stomatal conductance (g_s) and transpiration rate (E) in (a,c,e) *D. alatus* Roxb. ex G. Don and (b,d,f) *D. tuberculatus* Roxb. grown under well watering (WW) and water deficit (WD) conditions.

demonstrated (Fig. 2e,f).

In canopy layers of mono-plantation, C_i in *H. brasiliensis* and *E. camaldulensis* trees under water deficit condition was decreased according to increased light intensity, whereas it was unchanged under well watering condition (Fig. 3a,b). Inter-

estingly, C_i in *E. camaldulensis* under water deficit was better than well watering (Fig. 3b) whereas it was lower in *H. brasiliensis* (Fig. 3a). *E. camaldulensis* was identified as light sensitive based on the increase in P_n in response to high light intensity (Fig. 3d). In water-deficit stress, P_n in *H. brasiliensis*

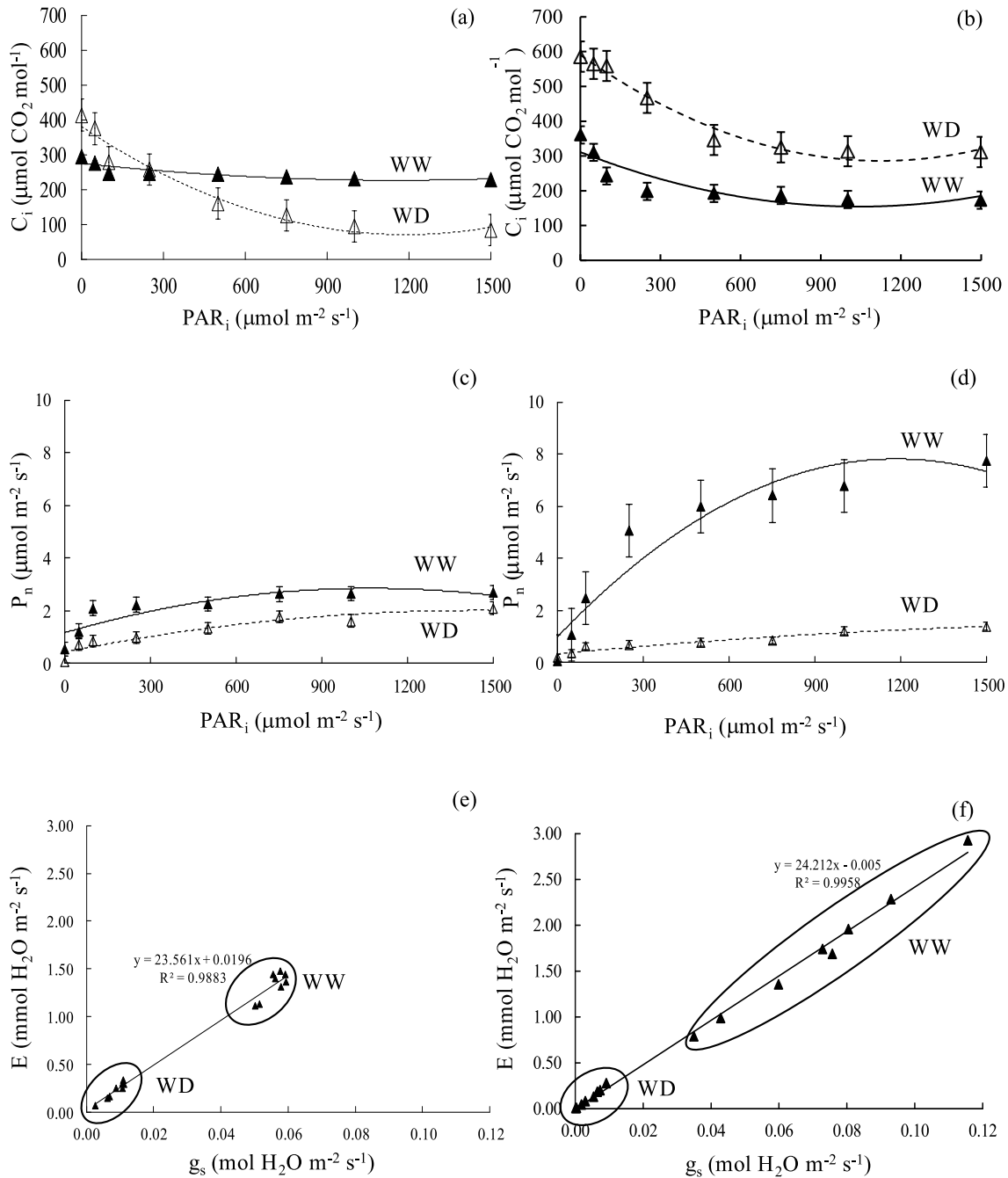


Fig. 3 Same as Fig. 2, for (a,c,e) *H. brasiliensis* Muell. Arg. and (b,d,f) *E. camaldulensis* Dehnh.

was maintained, but P_n of *E. camaldulensis* plant was very low in each light flux when compared with well-watered plants (Fig. 2c,d). Progressive relationships between g_s and E in *H. brasiliensis* ($R^2 = 0.988$) and *E. camaldulensis* ($R^2 = 0.996$) were demonstrated (Fig. 3e,f).

In ground cover layer, C_i in *Colocasia* species showed a similar trend of decrease in response

to high light intensity in both well watering and water limited conditions (Fig. 4a,b). The P_n for *C. esculenta* under WW and WD was unchanged at high light intensity. Similarly, P_n was increased with an increase in PAR_i (Fig. 4c,d). A positive relation between g_s and E in *C. esculenta* ($R^2 = 0.807$) and *C. gigantea* ($R^2 = 0.971$) was demonstrated (Fig. 4e,f).

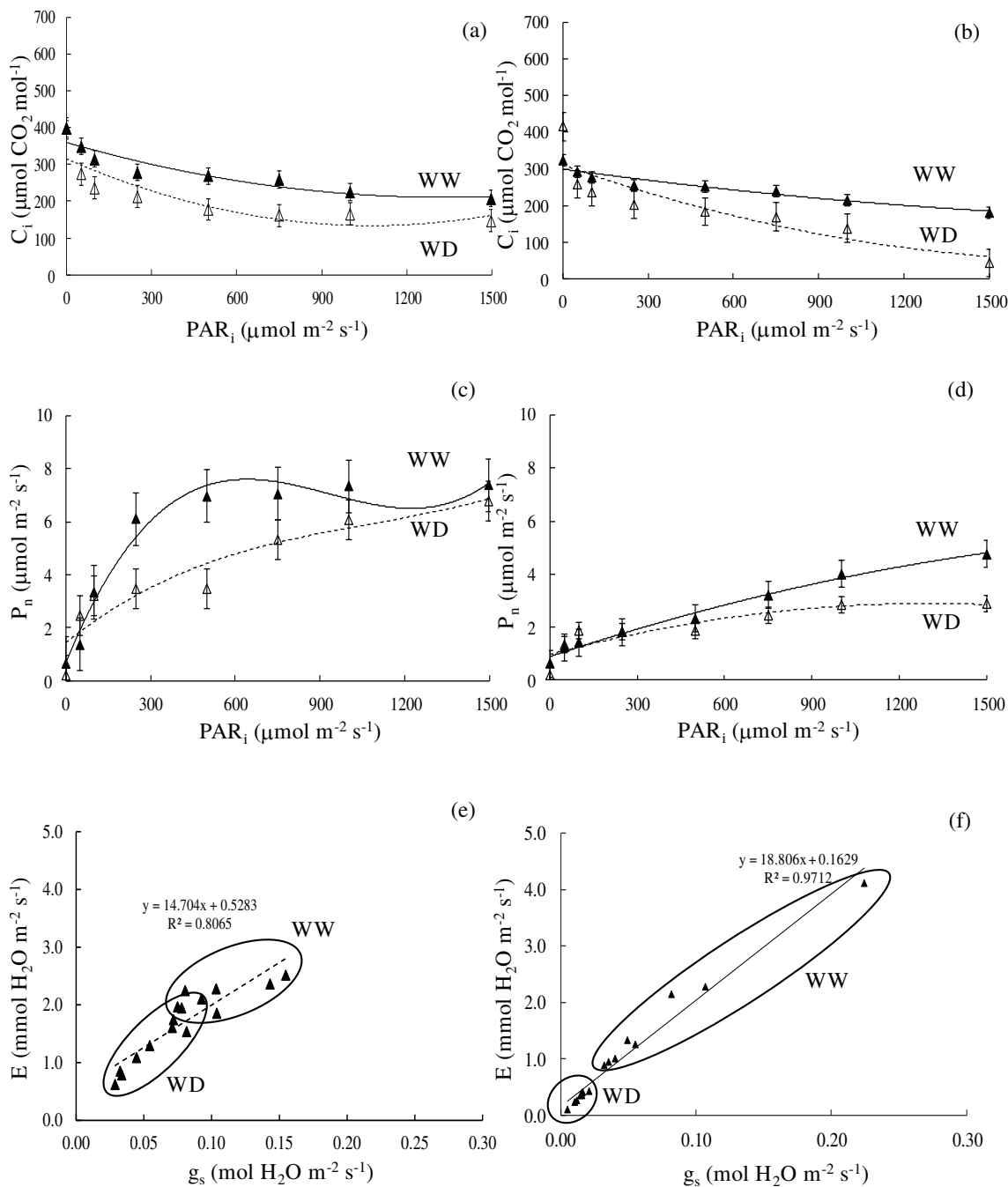


Fig. 4 Same as Fig. 2, for (a,c,e) *C. esculenta* (L.) Schott and (b,d,f) *C. gigantea* Hook. f.

DISCUSSION

In the present study, the plant injuries such as leaf chlorosis, leaf burn, plant wilting, and plant death depend on plant species and the water availability in the soil. Previously, the wilting state and mortality of plants under dry season in nine forest species have been categorized into two clusters:

cluster I (low survival rate with high wilting score or drought susceptible group) including *Beilschmiedia pendula*, *Calophyllum longifolium*, *Sorocea affinis* and *Tabebuia rosea*, and cluster II (high survival rate with low wilting score or drought sensitive group) including *Garcinia intermedia*, *Hybanthus prunifolius*, *Ouratea lucens*, *Pseudobombax septenatum* and *Psychotria horizontalis*²². In the present

study, stomatal density among treatments was unaffected, whereas the stomatal conductance in all plant species under water deficit condition was declined. In a study using 29 genotypes of poplar, total stomata density in plants grown under well-watered (757 stomata mm^{-2}) and water deficit conditions (769 stomata mm^{-2}) was unchanged, while stomatal conductance or stomatal closure in plants under water deficit declined by 28% when compared with well-watered conditions²⁶. In contrast, the stomatal density in olive changed under longer durations of water deficit^{16,18}.

In general, the stomatal closure relating to g_s and E reduction was evidently observed when plants were subjected to water deficit, resulting in limited CO_2 assimilation. In the present study, P_n , g_s , E , and WUE reduction in several plant species under water shortage situation was described. Similar reductions in P_n (74% of control), g_s (75% of control), and E (92%) were observed in plants in response to water-deficit stress (25% FC) in *Populus przewalskii*²⁷. In addition, rainy and dry seasons introduce a large gap of water availability in the soil, leading to retardation of photosynthetic abilities. For example, photosynthetic capacity (A_{max}) in *D. retusus*, *Hopea hainanensis*, *Parashorea chinensis*, and *Vatica xishuangbannaensis* under dry season was decreased by 52%, 52%, 58%, and 63%, respectively, when compared with the rainy season²⁸. A_{max} and g_s in *Miconia ciliata* are very low in the dry season, subsequently lifting up by irrigation and wet season²⁹. In addition, P_n and g_s in Bornean heath forest including *Cotylelobium burckii*, *D. borneensis*, *H. pentanervia*, *Shorea multiflora*, *Fordia splendidissima*, *Tristanopsis obovata*, and *Cleistanthus baramicus* in the dry season (December–April) are decreased³⁰. The decline in these parameters depends on plant species³¹ and the degree of drought stress³².

A decrease in C_i in response to PAR_i in each plant genotype was established. A rate of C_i reduction depending on the plant species and the degree of water deficit conditions was observed. The decrease in C_i of well-watered plants of *Jatropha curcas* cultivars in Indonesia and Cape Verde islands was confirmed in relation to increased PPF³¹. In general, C_i of well-watered plants was maintained at a higher level than in water stressed plants^{17,33}. In the dry season, C_i of controlled plants of *M. ciliata* was lower than that of irrigated plants and it increased in the rainy season²⁹. Interestingly, the C_i in water stressed *E. camaldulensis* was accumulated in the higher concentrations than that of well-watered plants. Leaf burn was evidently observed

in *E. camaldulensis* under water deficit, leading to high levels of intracellular CO_2 . Similar results have been reported in *Vitis vinifera* cultivars Khoshnave, Askan, and Bidane-Sefid that showed an increase in C_i when plants were subjected to the enhanced degree of water-deficit stresses³². In addition, C_i in *D. tuberculatus* was retained in both WW and WD conditions. In general, the C_i in *Podocarpus lawrencei* plants grown under drought condition fluctuates relating to CO_2 assimilation rate ($R^2 = 0.935$) and it fits to the linear curve in both watered and unwatered plants³⁴.

CO_2 assimilation rate or P_n in the leaf tissues of each plant species was increased in relation to the density of light flux. Furthermore, P_n in the plants grown under water deficit conditions was slightly increased and demonstrated a big gap between well-watered and water stressed plants, especially in *D. alatus*, *D. tuberculatus* and *E. camaldulensis*, which was evidently demonstrated by the visual symptoms such as wilting, leaf chlorosis and leaf burn, respectively. In addition, the P_n of four dipterocarp species, i.e., *D. retusus*, *H. hainanensis*, *P. chinensis*, and *V. xishuangbannaensis* under dry season was lower than that under rainy season²⁸. Based on P_n reduction with light response curve in *P. cathayana* when subjected to 30% FC, the reduction rate was more in female plants than in male plants³⁵, indicating higher sensitivity of female plants towards the drought conditions. In addition, P_n in water-deficit stress of *C. esculenta* was sustained nearby in unstressed plants. It possible that an enriched epicuticular wax in the abaxial leaves may play a key role as water loss prevention when subjected to water deficit³⁶. Likewise, there are some other factors in the light responses when plants were exposed to water deficit conditions, i.e., leaf position (exposed and shaded leaves)^{37,38}, and a strength of water-deficit stresses in *Eragrostis curvula*³⁹, *Quercus ilex*⁴⁰, *H. brasiliensis*⁴¹, *Lycium nodosum*⁴², and *Tamarix chinensis*⁴³. A reduction in g_s and E in each plant grown under water deficit condition was evidently established and subsequently a positive relation was observed^{44,45}. In addition, a positive relation between g_s and P_n in leaves of *L. nodosum* ($R^2 = 0.87$)⁴², *Vitis vinifera* ($R^2 = 0.83$)³² and *Symplocos racemosa* ($R^2 = 0.80$)⁴⁶ was demonstrated.

In conclusion, the toxic symptoms, i.e., wilting (in *D. alatus*), leaf chlorosis (in *D. tuberculatus*), and leaf burn (in *E. camaldulensis*) were evidently observed when the water availability in the soil was declined, leading to stomatal closure to prevent wa-

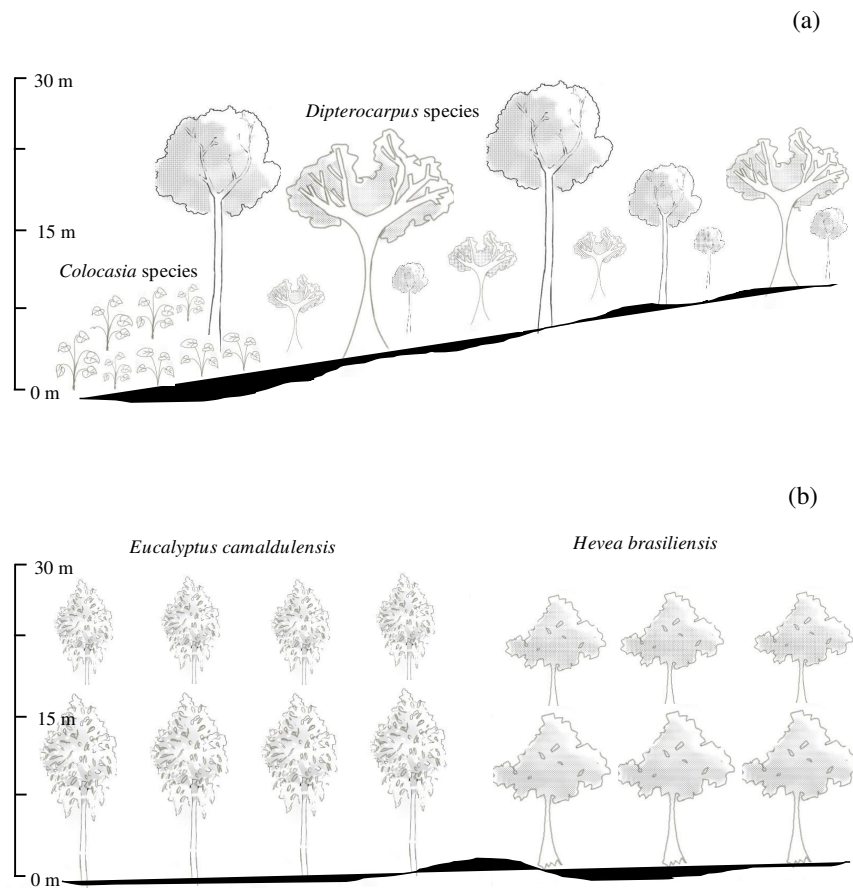


Fig. 5 (a) A scheme of mixed deciduous forest with *C. esculenta* (L.) Schott and *C. gigantea* Hook. f. grown under high humidity with shading light environments. (b) A scheme of monoculture farmed forest species including *E. camaldulensis* and *H. brasiliensis*.

ter loss and consequently reduced assimilation rate. Based on the photosynthetic response, eucalyptus demands more water supply and high light intensity when compared with rubber tree, which is generally grown as mono-plantation. In addition, the light response curve was a key indicator of water deficit conditions in case of emergent layer, canopy layer, and ground cover layer, and it should be managed while planning the mixed deciduous plantations and mono-plantation strategies as demonstrated in the models (Fig. 5).

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REFERENCES

1. Leakey RRB (1999) Potential for novel food products from agroforestry: a review. *Food Chem* **66**, 1–14.
2. Abbas F, Hammad HM, Fahad S, Cerdà A, Rizwan M, Farhad W, Ehsan S, Bakhat HF (2017) Agroforestry: a sustainable environmental practice for carbon sequestration under the climate change scenarios—a review. *Environ Sci Pollut Res* **24**, 11 177–91.
3. Luedeling E, Smethurst PJ, Baudron F, Bayala J, Huth NI, van Noordwijk M, Ong CK, Mulia R, Lusiana B, Muthuri C, Sinclair FL (2016) Field-scale modeling of tree-crop interactions: Challenges and development needs. *Agr Syst* **142**, 51–69.
4. Xuluc-Tolosa FJ, Vester HFM, Ramírez-Marcial N, Castellanos-Albores J, Lawrence D (2003) Leaf litter decomposition of tree species in three successional phases of tropical dry secondary forest in Campeche, Mexico. *Forest Ecol Manag* **174**, 401–12.
5. Muleta D, Assefa F, Nemomissa S, Granhall U (2008) Distribution of arbuscular mycorrhizal fungi spores in soils of smallholder agroforestry and monocultural coffee systems in southwestern Ethiopia. *Biol Fertil Soils* **44**, 653–9.
6. Sanou J, Bayala J, Bazié P, Teklehaimanot Z (2012)

- Photosynthesis and biomass production by millet (*Pennisetum glaucum*) and taro (*Colocasia esculenta*) grown under baobab (*Adansonia digitata*) and néré (*Parkia biglobosa*) in an agroforestry parkland system of Burkina Faso (West Africa). *Exp Agr* **48**, 283–300.
7. Kwesiga F, Akinnifesi FK, Mafongoya PL, McDermott MH, Agumya A (2003) Agroforestry research and development in southern Africa during the 1990s: Review and challenges ahead. *Agroforest Syst* **59**, 173–86.
 8. Bellow JG, Nair PKR (2003) Comparing common methods for assessing understory light availability in shaded-perennial agroforestry systems. *Agr Forest Meteorol* **114**, 197–211.
 9. Schaller M, Schroth G, Beer J, Jiménez F (2003) Species and site characteristics that permit the association of fast-growing tree with crops: The case of *Eucalyptus deglupta* as coffee shade in Costa Rica. *Forest Ecol Manag* **175**, 205–15.
 10. van Kanten R, Schroth G, Beer J, Jiménez F (2005) Fine-root dynamics of coffee in association with two shade trees in Costa Rica. *Agroforest Syst* **63**, 247–61.
 11. Stringer LC, Dyer JC, Reed MS, Dougill AJ, Twyman C, Mkwambisi D (2009) Adaptations to climate change, drought and desertification: local insights to enhance policy in southern Africa. *Environ Sci Pol* **12**, 748–65.
 12. Loewenstein NJ, Pallardy SG (1998) Drought tolerance, xylem sap abscisic acid and stomatal conductance during soil drying: a comparison of canopy trees of three temperate deciduous angiosperms. *Tree Physiol* **18**, 431–9.
 13. Aasamaa K, Sober A, Hartung W, Niinemets U (2002) Rates of stomatal opening, shoot hydraulic conductance and photosynthetic characteristics in relation to leaf abscisic acid concentration in six temperate deciduous trees. *Tree Physiol* **22**, 267–76.
 14. Neumann PM (2008) Coping mechanisms for crop plants in drought-prone environments. *Ann Bot* **101**, 901–7.
 15. Munné-Bosch S, Alegre L (2004) Die and let live: Leaf senescence contributes to plant survival under drought stress. *Funct Plant Biol* **31**, 203–16.
 16. Bosabalidis AM, Kofidis G (2002) Comparative effects of drought stress on leaf anatomy of two olive cultivars. *Plant Sci* **163**, 375–9.
 17. Brodribb T (1996) Dynamics of changing intercellular CO₂ concentration (C_i) during drought and determination of minimum functional C_i. *Plant Physiol* **111**, 179–85.
 18. Guerfel M, Baccouri O, Boujnah D, Chaïbi W, Zarrouk M (2009) Impacts of water stress on gas exchange, water relations, chlorophyll content and leaf structure in two main Tunisian olive (*Olea europaea* L.) cultivars. *Sci Hort* **119**, 257–63.
 19. Carter EB, Theodorou MK, Morris P (1997) Responses of *Lotus corniculatus* to environmental change. I. Effects of elevated CO₂, temperature and drought on growth and plant development. *New Phytol* **136**, 245–53.
 20. Engelbrecht BMJ, Kursar TA (2003) Comparative drought-resistance of seedlings of 28 species of co-occurring tropical woody plants. *Oecologia* **136**, 383–93.
 21. Comita LS, Engelbrecht BMJ (2009) Seasonal and spatial variation in water availability drive habitat associations in a tropical forest. *Ecology* **90**, 2755–65.
 22. Poorter L, Markesteijn L (2008) Seedling traits determine drought tolerance of tropical tree species. *Biotropica* **40**, 321–31.
 23. Baltzer JL, Davies SJ, Bunyavejchewin S, Noor NSM (2008) The role of desiccation tolerance in determining tree species distributions along the Malay-Thai Peninsula. *Funct Ecol* **22**, 221–31.
 24. Cha-um S, Kirdmanee C (2008) Assessment of salt tolerance in *Eucalyptus*, Rain tree and Thai neem under laboratory and the field conditions. *Pakistan J Bot* **40**, 2041–51.
 25. Aleric KM, Kirkman LK (2005) Growth and photosynthetic responses of the federally endangered shrub, *Lindera melissifolia* (Lauraceae), to varied light environments. *Am J Bot* **92**, 682–9.
 26. Monclus R, Dreyer E, Villar M, Delmotte FM, Delay D, Petit JM, Barbaroux C, Le Thiec D, Bréchet C, Brignolas F (2006) Impact of drought on productivity and water use efficiency in 29 genotypes of *Populus deltoides* × *Populus nigra*. *New Phytol* **169**, 765–77.
 27. Yin CY, Berninger F, Li CY (2006) Photosynthetic responses of *Populus przewalskii* subjected to drought stress. *Photosynthetica* **44**, 62–8.
 28. Zhang JL, Meng LZ, Cao KF (2008) Sustained diurnal photosynthetic depression in uppermost-canopy leaves of four dipterocarp species in the rainy and dry seasons: does photorespiration play a role in photoprotection? *Tree Physiol* **29**, 217–28.
 29. Fortini LB, Mulkey SS, Zarin DJ, Vasconcelos SS, de Carvalho CJR (2003) Drought constraints on leaf gas exchange by *Miconia ciliata* (Melastomataceae) in the understory of an eastern Amazonian regrowth forest stand. *Am J Bot* **90**, 1064–70.
 30. Cao KF (2000) Water relations and gas exchange of tropical samplings during a prolonged drought in a Bornean heath forest, with reference to root architecture. *J Trop Ecol* **16**, 101–16.
 31. Sapeta H, Costa JM, Lourenço T, Maroco J, van der Linde P, Oliveira MM (2013) Drought stress response in *Jatropha curcas*: Growth and physiology. *Environ Exp Bot* **85**, 76–84.
 32. Ghaderi N, Talaie AR, Ebadi A, Lessani H (2011) The physiological response of three Iranian grape cultivars to progressive drought stress. *J Agr Sci Tech* **13**, 601–10.
 33. Souza RP, Machado EC, Silva JAB, Lagôa AMMA, Silveira JAG (2004) Photosynthetic gas exchange,

- chlorophyll fluorescence and some associated metabolic changes in cowpea (*Vigna unguiculata*) during water stress and recovery. *Environ Exp Bot* **51**, 45–56.
34. Centritto M (2005) Photosynthetic limitations and carbon partitioning in cherry in response to water deficit and elevated [CO₂]. *Agr Ecosyst Environ* **106**, 233–42.
 35. Xu X, Peng G, Wu C, Korpelainen H, Li C (2008) Drought inhibits photosynthetic capacity more in females than in male of *Populus cathayana*. *Tree Physiol* **28**, 1751–9.
 36. Kim KS, Park SH, Jenks MA (2007) Changes in leaf cuticular waxes of sesame (*Sesamum indicum* L.) plants exposed to water deficit. *J Plant Physiol* **164**, 1134–43.
 37. Muraoka H, Tang Y, Koizumi H, Washitani I (2002) Effects of light and soil water availability on leaf photosynthesis and growth of *Arisaema heterophyllum*, a riparian forest understory plant. *J Plant Res* **115**, 419–27.
 38. Sofo A, Dichio B, Montanaro G, Xiloyannis C (2009) Photosynthetic performance and light response of two olive cultivars under different water and light regimes. *Photosynthetica* **47**, 602–8.
 39. Colom MR, Vazzana C (2001) Drought stress effects on three cultivars of *Eragrostis curvula*: Photosynthesis and water relation. *Plant Growth Regul* **34**, 195–202.
 40. Peña-Rojas K, Aranda X, Fleck I (2004) Stomatal limitation to CO₂ assimilation and down-regulation of photosynthesis in *Quercus ilex* resprouts in response to slowly imposed drought. *Tree Physiol* **24**, 813–22.
 41. Sangsing K, Le Roux X, Kasemsap B, Thanisawan-yangkura S, Sangkhasila K, Gohet E, Thaler P (2004) Photosynthetic capacity and effect of drought on leaf gas exchange in two rubber (*Hevea brasiliensis*) clones. *Kasetsart J (Nat Sci)* **38**, 111–22.
 42. Tezara W, Martínez D, Rengifo E, Herrera A (2003) Photosynthetic responses of the tropical spiny shrub *Lycium nodosum* (Solanaceae) to drought, soil salinity and saline spray. *Ann Bot* **92**, 757–65.
 43. Xia JB, Zhao ZG, Sun JK, Liu JT, Zhao YY (2017) Response of stem sap flow and leaf photosynthesis in *Tamarix chinensis* to soil moisture in the Yellow River Delta, China. *Photosynthetica* **55**, 368–77.
 44. Jarvis PG, McNaughton KG (1986) Stomatal control of transpiration: scaling up from leaf to region. *Adv Ecol Res* **15**, 1–49.
 45. Mäkelä A, Berninger F, Hari P (1996) Optimal control of gas exchange during drought: Theoretical analysis. *Ann Bot* **77**, 461–8.
 46. Zhang JL, Zhu JJ, Cao KF (2007) Seasonal variation in photosynthesis in six woody species with different leaf phenology in a valley savanna in southwestern China. *Trees* **21**, 631–43.